

10/018449 DEC 2001

FORM PTO-1390 (Modified) (REV 11-2000)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER <b>10848-016001</b>	
<b>TRANSMITTAL LETTER TO THE UNITED STATES</b> <b>DESIGNATED/ELECTED OFFICE (DO/EO/US)</b> <b>CONCERNING A FILING UNDER 35 U.S.C. 371</b>				U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR) <b>10/018449</b>	
INTERNATIONAL APPLICATION NO <b>PCT/DE00/01945</b>		INTERNATIONAL FILING DATE <b>June 10, 2000</b>		PRIORITY DATE CLAIMED <b>June 14, 1999</b>	
TITLE OF INVENTION <b>METHOD AND DEVICE FOR IDENTIFYING A POLYMER</b>					
APPLICANT(S) FOR DO/EO/US <b>Wolf Bertling et al.</b>					
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:					
<ol style="list-style-type: none"> <li>1. <input checked="" type="checkbox"/> This is a <b>FIRST</b> submission of items concerning a filing under 35 U.S.C. 371.</li> <li>2. <input type="checkbox"/> This is a <b>SECOND</b> or <b>SUBSEQUENT</b> submission of items concerning a filing under 35 U.S.C. 371.</li> <li>3. <input type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.</li> <li>4. <input type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31).</li> <li>5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371 (c) (2))           <ol style="list-style-type: none"> <li>a. <input type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau).</li> <li>b. <input checked="" type="checkbox"/> has been communicated by the International Bureau.</li> <li>c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US).</li> </ol> </li> <li>6. <input checked="" type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).           <ol style="list-style-type: none"> <li>a. <input checked="" type="checkbox"/> is attached hereto.</li> <li>b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4).</li> </ol> </li> <li>7. <input type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))           <ol style="list-style-type: none"> <li>a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau).</li> <li>b. <input type="checkbox"/> have been communicated by the International Bureau.</li> <li>c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired.</li> <li>d. <input type="checkbox"/> have not been made and will not be made.</li> </ol> </li> <li>8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).</li> <li>9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).</li> <li>10. <input type="checkbox"/> An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).</li> <li>11. <input type="checkbox"/> A copy of the International Preliminary Examination Report (PCT/IPEA/409).</li> <li>12. <input checked="" type="checkbox"/> A copy of the International Search Report (PCT/ISA/210).</li> </ol>					
<b>Items 13 to 20 below concern document(s) or information included:</b>					
<ol style="list-style-type: none"> <li>13. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98.</li> <li>14. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.</li> <li>15. <input checked="" type="checkbox"/> A <b>FIRST</b> preliminary amendment.</li> <li>16. <input type="checkbox"/> A <b>SECOND</b> or <b>SUBSEQUENT</b> preliminary amendment.</li> <li>17. <input type="checkbox"/> A substitute specification.</li> <li>18. <input type="checkbox"/> A change of power of attorney and/or address letter.</li> <li>19. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.</li> <li>20. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4).</li> <li>21. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).</li> <li>22. <input checked="" type="checkbox"/> Certificate of Mailing by Express Mail</li> <li>23. <input checked="" type="checkbox"/> Other items or information:</li> </ol>					
<b>Verification of Translation (1 pg.); References cited in Search Report (7 references); postcard and check</b>				"EXPRESS MAIL" Mailing Label Number <u>EL5896418711US</u> Date of Deposit <u>December 14, 2001</u> I hereby certify under 37 CFR 1.10 that this correspondence is being deposited with the United States Postal Service as "Express Mail Post Office To Addressee" with sufficient postage on the date indicated above and is addressed to the Commissioner of Patents and Trademarks, Washington, D.C. 20231. <i>Josh Hattinen</i>	

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.53(b)(2))		INTERNATIONAL APPLICATION NO.		ATTORNEY'S DOCKET NUMBER	
10/018449		PCT/DE00/01945		10848-016001	
24. The following fees are submitted:.				CALCULATIONS PTO USE ONLY	
BASIC NATIONAL FEE ( 37 CFR 1.492 (a) (1) - (5) ) :					
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO . . . . .				\$1000.00	
<input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO . . . . .				\$860.00	
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO . . . . .				\$710.00	
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) . . . . .				\$690.00	
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) . . . . .				\$100.00	
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$860.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	32 - 20 =	12	x \$18.00	\$216.00	
Independent claims	2 - 3 =	0	x \$80.00	\$0.00	
Multiple Dependent Claims (check if applicable).			<input type="checkbox"/>	\$0.00	
TOTAL OF ABOVE CALCULATIONS =				\$1,076.00	
<input checked="" type="checkbox"/> Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.				\$538.00	
SUBTOTAL =				\$538.00	
Processing fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30 +				\$0.00	
TOTAL NATIONAL FEE =				\$538.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).			<input type="checkbox"/>	\$0.00	
TOTAL FEES ENCLOSED =				\$538.00	
				Amount to be refunded	\$
				charged	\$
a. <input checked="" type="checkbox"/> A check in the amount of \$538.00 to cover the above fees is enclosed.					
b. <input type="checkbox"/> Please charge my Deposit Account No. in the amount of to cover the above fees. A duplicate copy of this sheet is enclosed.					
c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 06-1050 A duplicate copy of this sheet is enclosed.					
d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO:					
Mark S. Ellinger Fish & Richardson P.C., P.A. 60 South Sixth Street, Suite 3300 Minneapolis, Minnesota 55402			M. Angela Parsons SIGNATURE M. Angela Parsons NAME 44,282 REGISTRATION NUMBER December 14, 2001 DATE		

531 Rec'd PCT: 14 DEC 2001

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Wolf Bertling et al.                      Art Unit : Unknown  
Serial No. :    Examiner : Unknown  
Filed : 14 December 2001  
Title : Method and Device For Identifying a Polymer

Commissioner for Patents  
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Prior to examination, please amend the application as follows:

In the Specification:

Please delete the paragraph on page 2, lines 6-8.

In the Claims:

Please cancel claim 4.

Please amend claims 2-7, 10, 12-13, 17-20, 22-24, and 27. A complete set of pending claims, including those amended herein, is as follows:

1. A method for identifying a first polymer (4, 7) which is bound to a first phase (5) which reflects electromagnetic waves, which method has the following steps:
  - a) bringing the first polymer (4, 7) into contact to a second polymer (3, 8) which has affinity to the first polymer (4, 7) and which is bound, by way of metallic clusters (2), to a solid second phase (1) which is pervious to electromagnetic waves,
  - b) irradiating the second phase (1) with electromagnetic waves, and
  - c) detecting the change in the properties of the reflected electromagnetic waves.
2. (Amended) The method as claimed in claim 1, characterized in that light is used as the electromagnetic waves.

## CERTIFICATE OF MAILING BY EXPRESS MAIL

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December 14, 2001

Date of Deposit

Signature

Josh Halttunen

Typed or Printed Name of Person Signing Certificate

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3. (Amended) The method as claimed in claim 1, characterized in that the property change which is measured is the absorption in a predetermined spectrum before and/or after the first polymer (4, 7) has been brought into contact with the second polymer (3, 8).

5. (Amended) The method as claimed in claim 1, characterized in that the property change which is measured is the chronological change in the absorption and/or reflection when the first polymer (4, 7) and the second polymer (3, 8) are brought into contact and/or separated.

6. (Amended) The method as claimed in claim 1, characterized in that the property change is measured at several angles of incidence which differ from each other.

7. (Amended) The method as claimed in claim 1, characterized in that the metallic clusters (2) are evaporation-coated directly onto the second phase (1) or are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).

8. The method as claimed in claim 7, characterized in that a layer formed from the second polymer (3, 8) is applied to the surface of the second phase (1).

9. The method as claimed in claim 8, characterized in that at least one layer formed from the first polymer (4, 7) is intercalated between the layer which is applied to the surface and the layer which is bonded to the metallic clusters (2).

10. (Amended) The method as claimed in claim 1, characterized in that a layer formed from the first polymer (4, 7) is applied to the surface of the first phase (5).

11. The method as claimed in claim 10, characterized in that a layer sequence formed from the first polymer (4, 7) and the second polymer (3, 8) is applied to the surface.

12. (Amended) The method as claimed in claim 1, characterized in that the first polymer (4, 7) and/or second polymer (3, 8) employ and/or employs DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptide nucleic acid (PNA) or a ligand thereof, or polyacrylic acid, polyethylenimine or poly(D-glucosamine).

13. (Amended) The method as claimed in claim 1, characterized in that, in the step denoted with the letter a, at least one other polymer, which is bound to the first phase (5), is brought into contact with the second polymer (3, 8).

14. The method as claimed in claim 13, characterized in that the polymers (3, 4, 7, 8) are applied to the first phase (5) in the form of a barcode.

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15. A device for identifying a first polymer (4, 7) which is bound to a first phase (5) which reflects electromagnetic waves, characterized in that a second polymer (3, 8), which has affinity to the first polymer (4, 7), is bound, by way of metallic clusters (2), to the surface of a second phase (1) which is pervious to electromagnetic waves.

16. The device as claimed in claim 15, characterized in that the metallic clusters (2) are formed from silver, gold, aluminum, copper or indium.

17. (Amended) The device as claimed in claim 15, characterized in that the electromagnetic waves are light.

18. (Amended) The device as claimed in claim 15, characterized in that the second phase (1) is produced from a transparent material.

19. (Amended) The device as claimed in claim 15, characterized in that the first polymer (4, 7) and/or the second polymer (3, 8) is/are DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptidenucleic acid (PNA) or a ligand thereof, or polyacrylic acid, poly(D-glucosamine) or polyethylenimine.

20. (Amended) The device as claimed in claim 15, characterized in that a contrivance for determining the optical property of the reflected light is provided.

21. The device as claimed in claim 20, characterized in that the contrivance can be used to measure the absorption in a predetermined spectrum before and/or after the first polymer (4, 7) and the second polymer (3, 8) are brought into contact.

22. (Amended) The device as claimed in claim 20, characterized in that the contrivance can be used to measure the spectral shift of the reflective light.

23. (Amended) The device as claimed in claim 20, characterized in that the contrivance can be used to measure the optical property at several angles of incidence which differ from each other.

24. (Amended) The device as claimed in claim 15, characterized in that the metallic clusters (2) are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).

25. The device as claimed in claim 24, characterized in that a layer which is formed from the second polymer (3, 8) is applied to the surface on the second phase (1).

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26. The device as claimed in claim 25, characterized in that at least one layer which is formed from the first polymer (4, 7) is intercalated between the layer provided on the surface and the layer which is bonded to the metallic clusters (2).

27. (Amended) The device as claimed in claim 15, characterized in that a layer which is formed from the first polymer (4, 7) is applied to the surface of the first phase (5).

28. The device as claimed in claim 27, characterized in that a layer which is formed for the second polymer (3, 8) is applied to the layer which is provided on the surface.

Please add the following new claims:

- 29. The method as claimed in claim 2, wherein said light is LASER light.
- 30. The method as claimed in claim 2, wherein said light is monochromatic light.
- 31. The method as claimed in claim 30, wherein said property change which is measured is the spectral shift of said monochromatic light.
- 32. The device as claimed in claim 17, wherein said light is LASER light.
- 33. The device as claimed in claim 18, wherein said transparent material is plastic or glass.

In the Abstract:

Please add the Abstract on the attached page to the specification after the claims.

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### REMARKS

Applicants respectfully request entry of the amendments and remarks submitted herein. Claims 2-7, 10, 12, 13, 17-20, 22-24, and 27 have been amended to remove multiple claim dependencies, claim 4 has been canceled and new claims 29-33 have been added. Support for the claim amendments and new claims 29-33 can be found in the originally filed claims and throughout the specification. Therefore, claims 1-3 and 5-33 are currently pending. Attached is a marked-up version of the changes being made by the current amendments. Reconsideration of the pending application is respectfully requested.

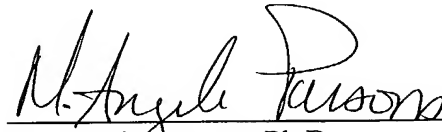
In addition, Applicants amended the specification to remove the paragraph on page 2 that refers to claim numbers, and to add an Abstract. The attached Abstract is the Abstract that was published with the PCT application. Therefore, Applicants submit that there is no new matter introduced by these amendments.

### CONCLUSION

Applicants ask that claims 1-3 and 5-33 be examined. The enclosed filing fee takes into account the new claims added by this Preliminary Amendment. Please apply any other charges or credits to Deposit Account No. 06-1050.

Respectfully submitted,

Date: December 14, 2001

  
\_\_\_\_\_  
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VERSION WITH MARKINGS TO SHOW CHANGES MADE

In the Specification:

The paragraph on page 2, lines 6-8 has been deleted.

In the Claims:

Please cancel claim 4.

Claims 2-7, 10, 12-13, 17-20, 22-24, and 27 have been amended as follows:

2. (Amended) The method as claimed in claim 1, characterized in that light[, preferably LASER light,] is used as the electromagnetic waves.

3. (Amended) The method as claimed in claim 1 [either of the preceding claims], characterized in that the property change which is measured is the absorption in a predetermined spectrum before and/or after the first polymer (4, 7) has been brought into contact with the second polymer (3, 8).

5. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that the property change which is measured is the chronological change in the absorption and/or reflection when the first polymer (4, 7) and the second polymer (3, 8) are brought into contact and/or separated.

6. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that the property change is measured at several angles of incidence which differ from each other.

7. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that the metallic clusters (2) are evaporation-coated directly onto the second phase (1) or are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).

10. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that a layer formed from the first polymer (4, 7) is applied to the surface of the first phase (5).

12. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that the first polymer (4, 7) and/or second polymer (3, 8) employ and/or employs



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DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptide nucleic acid (PNA) or a ligand thereof, or polyacrylic acid, polyethylenimine or poly(D-glucosamine).

13. (Amended) The method as claimed in claim 1 [one of the preceding claims], characterized in that, in the step denoted with the letter a, at least one other polymer, which is bound to the first phase (5), is brought into contact with the second polymer (3, 8).

17. (Amended) The device as claimed in claim 15 [or 16], characterized in that the electromagnetic waves are light[, preferably LASER light].

18. (Amended) The device as claimed in claim [one of claims] 15 [to 17], characterized in that the second phase (1) is produced from a transparent material[, such as plastic or glass].

19. (Amended) The device as claimed in claim [one of claims] 15 [to 18], characterized in that the first polymer (4, 7) and/or the second polymer (3,8) is/are DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptidenucleic acid (PNA) or a ligand thereof, or polyacrylic acid, poly(D-glucosamine) or polyethylenimine.

20. (Amended) The device as claimed in claim [one of claims] 15 [to 19], characterized in that a contrivance for determining the optical property of the reflected light is provided.

22. (Amended) The device as claimed in claim 20 [or 21], characterized in that the contrivance can be used to measure the spectral shift of the reflective light.

23. (Amended) The device as claimed in claim 20 [one of claims 15 to 23], characterized in that the contrivance can be used to measure the optical property at several angles of incidence which differ from each other.

24. (Amended) The device as claimed in claim [one of claims] 15 [to 23], characterized in that the metallic clusters (2) are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).

27. (Amended) The device as claimed in claim [one of claims] 15 [to 26], characterized in that a layer which is formed from the first polymer (4, 7) is applied to the surface of the first phase (5).

New claims 29-33 have been added.

In the Abstract:

The Abstract on the attached page has been added to the application.

**ABSTRACT OF THE DISCLOSURE**

The invention relates to a method for identifying a first polymer (4, 7) which is bonded to a first phase (5) that reflects electromagnetic waves. Said method comprises the following steps: (a) bringing the first polymer (4, 7) into contact with a second polymer (3, 8), which is bonded to a solid second phase (1) by metallic clusters (2), said second phase being permeable to electromagnetic waves; (b) irradiating the second phase (1) with electromagnetic waves; and (c) determining the change in the properties of the reflected electromagnetic waves.

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## Description

## Method and device for identifying a polymer

5

The invention relates to a method and a device for identifying a first polymer which is bound to a first phase which reflects electromagnetic waves.

- 10 WO 98/48275 discloses an optical sensor which can be used for detecting nucleic acids and proteins and their ligands. For the detection, the optical sensor is, for example, dipped into a nucleic acid-containing solution. After the sensor has been rinsed and dried,
- 15 its optical property can then be determined. The method using the known sensor requires several steps; it is time-consuming.

- WO 97/04129 discloses a method for detecting nucleic acid sequences; in this method, a first nucleic acid sequence is immobilized on a solid surface. The hybridization with a second complementary nucleic acid sequence is detected using surface-sensitive detection methods under specific conditions.

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- WO 91/02981 describes a method for detecting an analyte using surface plasmon resonance spectroscopy. In this case too, an analyte is immobilized on a metal surface. For the detection, the analyte has in turn to be
- 30 brought into contact with a solution.

- In addition, US 5,485,277 discloses a sensor for carrying out surface plasmon resonance spectroscopy. The sensor possesses a planar wave conductor which
- 35 possesses a large number of reflector surfaces.

The object of the invention is to eliminate the disadvantages of the prior art. The intention is, in

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particular, to specify a method and a device which can be used to rapidly and readily detect polymers, in particular biochemical molecules, which are bound to a solid phase.

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This object is achieved by means of the features in claims 1 and 15. Expedient refinements of the invention ensue from the features in claims 2 to 14 and 16 to 28.

10 In accordance with the invention, a method is provided for identifying a first polymer which is bound to a first phase which reflects electromagnetic waves, which method has the following steps:

15 a) bringing the first polymer into contact with a second polymer which has affinity for the first polymer and which is bound, by way of metallic clusters, to a solid second phase which is pervious to electromagnetic waves,

20

b) irradiating the second phase with electromagnetic waves, and

c) detecting the change in the properties of the  
25 reflected electromagnetic waves.

According to the method according to the invention, the polymer to be detected, for example a biochemical molecule, does not necessarily have to be present in  
30 solution. It can, for example, also be bound, for labeling purposes, to a solid body, such as a banknote. By simply bringing into contact the second electromagnetic wave-pervious phase and measuring the optical properties of the reflected light, it is  
35 possible to determine immediately whether the polymer which is to be detected is bound to the first solid phase. The method can be carried out rapidly and simply.

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Advantageously, the electromagnetic waves employed consist of light, preferably LASER light. The properties of reflected light can be determined in a particularly simple manner.

The property change which is measured can be the absorption in a predetermined spectrum before and/or after the first and second polymers have been brought into contact. Furthermore, when monochromatic light is used, the property change which is measured can be the spectral shift. Furthermore, the property change which is measured can be the chronological change in the absorption and/or reflection when the first and second polymers are brought into contact and/or separated. The property change can be measured under several angles of incidence which differ from each other. It is also possible to conceive of measuring other changes in the properties of the reflected light. The choice of which change is detected depends on the given circumstances.

The metallic clusters can either be evaporation-coated directly onto the second phase or else be bound to the second phase by way of a layer which is formed from the second polymer. A layer which is formed from the second polymer can be applied to the surface of the second phase. The second polymer can, for example, be poly(D-glucosamine). At least one layer which is formed from the first polymer can be intercalated between the layer which is applied to the surface and the layer which is bonded to the metallic clusters. A layer sequence which is formed from the first and the second polymers can also be intercalated. The provision of such a layer sequence contributes to the formation of signals which can be identified particularly clearly and rapidly. A layer formed from the first polymer can be applied to the surface of the first phase. The surface can be formed from an oxide layer of the metal, for example

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from an aluminum oxide layer. A layer sequence formed from the first and second polymers can also be provided on the surface, with it being possible for the outermost layer and the layer which is bound to the surface to be formed from the first polymer. The first polymer can, for example, be polyacrylic acid (= PAA).

A polynucleotide molecule such as DNA, RNA, ssDNA, ssRNA or synthetic analogs thereof, protein, peptide, peptide nucleic acid (PNA) or a ligand thereof, or polyacrylic acid, poly(D-glucosamine) or polyethylenimine, is expediently used as the first and/or second polymer. In principle, all biochemical molecules having recombinant properties are particularly suitable.

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In the step designated with the letter a, at least one further polymer which is bound to the first phase can also be brought into contact with the second polymer. This makes it possible to carry out a plurality of identification experiments simultaneously. Thus, the polymers can be used, for example, to form a barcode or a similar pattern on the surface of, for example, the first phase.

According to the invention, it is provided, in a device for identifying a first polymer which is bound to a first phase which reflects electromagnetic waves, that a second polymer, which has affinity for the first polymer, is bound, by way of metallic clusters, to the surface of a second phase which is pervious for electromagnetic waves.

The device according to the invention enables a first polymer to be identified rapidly and simply. There is no need to rinse and dry the device in order to measure the optical properties of the electromagnetic waves employed. Affinity is understood as meaning that the polymers can, by means of interactions, assume a bound

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or associated state. Such bonds can, for example, be hydrogen bonds, ionic bonds, hydrophobic bonds or covalent bonds. Other suitable bonds are complex bonds or bonds which are elicited by stearic effects. For  
5 example, the strains which are complementary to each other and which are present in biomolecules, such as DNA, are regarded as having affinity to each other; they are able to hybridize.

10 It has proved to be expedient to form the metallic clusters from silver, gold, aluminum, copper or indium. Polymers bind particularly well to these metals.

It is possible to use light, preferably LASER light, as  
15 the electromagnetic waves. Advantageously, the second phase is produced from a transparent material, such as plastic or glass. The first and/or second polymer can be DNA, RNA, protein, peptide, peptidenucleic acid or a ligand thereof, or polyacrylic acid, poly(D-  
20 glucosamine) or polyethylenimine. However, the polymer employed can also be ssDNA or ssRNA or synthetic analogs thereof.

A contrivance for determining the optical properties of  
25 the reflected light can be provided as an additional component of the device. The contrivance can be used to measure the absorption in a predetermined spectrum before and/or after the first and the second polymers have been brought into contact. In addition, the  
30 contrivance can be used to measure the spectral shift of the reflected light.

Expediently, the contrivance can be used to measure the  
optical property at several angles of incidence which  
35 differ from each other.

The metallic clusters can be bound to the second phase by way of a layer which is formed from the second

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polymer. Furthermore, a layer formed from the second polymer can be applied to the surface of the second phase. Expediently, at least one layer which is formed from the first polymer is also intercalated between the layer which is provided on the surface and the layer which is bound to the metallic clusters. A layer which is formed from the first polymer can be applied to the surface of the first phase and/or a layer which is formed from the second polymer can be applied to the layer which is provided on the surface. The layer complex which is described makes labeling possible and also makes it possible to identify the label simply and rapidly.

The invention is explained in more detail below with the aid of the exemplary embodiment which is depicted in the drawing. In the drawing:

Fig. 1 shows a diagrammatic view of a device,

Fig. 2 shows the device according to Fig. 1 in the non-hybridized case,

Fig. 3 shows the device according to Fig. 1 in the hybridized case,

Fig. 4 shows a diagrammatic view of another device, and

fig. 5 shows the detection of a label.

In Figs. 1-3, a second solid phase is produced, for example, from a glass support 1. Metallic clusters 2, for example gold clusters, are present on one surface of the glass support 1. A single-stranded DNA 3 is bound, as the second polymer, to the clusters 2. Another single-stranded DNA 4 is bound, as the first polymer, to a metal strip 5. The metal strip 5 can, in



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turn, be fixed, for example for labeling purposes, to banknotes (not depicted here).

5 Provided the DNA 3 and the other DNA 4 are brought into contact, two cases can be distinguished:

10 In the first case, which is shown in Fig. 2, the DNA 3 is not complementary to the other DNA 4. No hybridization takes place. A first distance  $d_1$  arises between the layer formed by the clusters 2 and the metal strip 5.

15 In the second case, which is shown in Fig. 3, the DNA 3 is complementary to the other DNA 4. The DNA 3 and the other DNA 4 hybridize. A smaller, second distance  $d_2$  arises between the layer formed by the clusters 2 and the metal strip 5.

20 A laser beam (not depicted here) which is incident through the glass support 1 is reflected at the [lacuna] by the metal strip 5. The properties of the reflected light depend on the distance  $d_1$  or  $d_2$  of the layer formed by the clusters 2 from the metal strip 5. Thus, the absorption, for example, is altered. By  
25 measuring the absorption, it is then possible to determine, in a simple manner, whether a hybridization is present or not. This makes it possible to identify the first polymer 4.

30 In order to produce the optical probe shown in Figs. 1-3, a glass support 1 is sputtered with gold. For this, the glass support 1 is suspended in a vacuum chamber in which gold foil is placed at the same time. After the vacuum chamber has been pumped down to  $10^{-2}$  mbar and  
35 subsequently flushed with argon gas, the pressure in the vacuum chamber is adjusted to about  $10^{-1}$  mbar. After that, a plasma is ignited, resulting in gold atoms being sputtered out of the foil. The gold atoms become

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deposited on the surface of the glass support 1. At a sputter current of 40 mA, a gold film having a mass thickness of 5 nm is formed after about 10 seconds. After that, the gold film is heated at about 200°C.

5 Round gold clusters 2, which are suitable for the desired color intensification effect, are formed. Subsequently, the glass support 1, which is coated with gold clusters 2, is dipped into a solution which contains oligonucleotides 3 which are provided with a

10 thiol group at their 5' ends. The oligonucleotides 3 settle on the gold clusters with the formation of a thiol bond.

An aluminum substrate 5 is, for example, used for producing the sample designated by the reference numbers 4 and 5. Using a defined electrochemical oxidation of the aluminum substrate 5, for example in 5% oxalic acid at 300mA and about 50 V, oxide layers of different thicknesses are generated by immersing the

20 aluminum substrate 5 more deeply, in a stepwise manner, into the solution. As a consequence of interference effects, these oxide layers have different colors.

In order to covalently couple other oligonucleotides 4, which are provided with amino groups at their 5' ends, the oxide layers are coated with a layer which carries a free amino group. For this, the aluminum substrate 5 is immersed, for about 30 minutes, in an approximately 10% aqueous solution of aminopropyl triethoxysilane at

30 a pH of about 9. The aluminum substrate is subsequently washed with water and dried for one hour in a drying oven at about 80°C.

The silanized aluminum substrates 5 which are produced in this way are then incubated for 12 hours in a 2.5% solution of glutardialdehyde which contains 50 mmol of NaCNBH<sub>3</sub>/l. After that, they are washed thoroughly with water.

In order to bind on the other oligonucleotide 4, the aluminum substrates 5 are incubated overnight, at 4°C, in an aqueous buffer solution which contains other  
5 oligonucleotides 4 at a concentration of 1  $\mu\text{mol/l}$ , 0.1  $\times$  PBS buffer and 50 mmol of  $\text{NaCNB}_3/\text{l}$ . After that, the aluminum substrates 5 are thoroughly rinsed once again with water. They then possess covalently bonded other oligonucleotides 4 on their surface.

10 With regard to other details, in particular the size of the cluster 2 and the distance parameters, the reader is referred to WO 98/48275, the disclosure content of which is hereby incorporated by reference.

In order to detect a hybridization between the DNA 3, or the detector oligonucleotide, and the other DNA 4, or the labeling oligonucleotide, the glass support 1 and the metal strip 5, e.g. the aluminum substrate which has been prepared by the above-described method, are pressed onto each other. As a consequence of different distances between the reflecting aluminum surface and the gold clusters 2, it is possible to recognize characteristic color patterns. These make it possible to conclude whether there is or is not hybridization between the labeling oligonucleotide and the detector oligonucleotide.

Fig. 4 shows another exemplary embodiment of the invention. An aluminum oxide layer 6 is located on a metal strip 5 prepared from aluminum. The aluminum oxide layer 6 is covered by a PAA layer 7. On top of this there then comes a poly(D-glucosamine) layer 8. Several such layer sequences consisting of PAA and poly(D-glucosamine) are provided. The uppermost layer is formed by a PAA layer 7.

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The probe consists of a glass support 1 which forms the substrate. A poly(D-glucosamine) layer 8 is provided on its surface. On top of that lies a PAA layer 7. Several layer sequences consisting of poly(D-glucosamine) 8 and PAA 7 are provided. Gold clusters 2 are bonded on a poly(D-glucosamine) layer 8 in the vicinity of the surface. On top of this there lies a further poly(D-glucosamine) layer 8. For test purposes, a section of this layer is overlaid with a PAA layer 7.

10

In order to produce the coated aluminum strip 5, the latter is coated by immersing it alternately in a polyacrylic acid-containing solution and in a poly(D-glucosamine) solution. In each case, the coating time was 15 minutes and the concentration of the solutions was 0.5 g/l. For coating with the gold clusters 2, the probe is immersed in a solution which contains approximately 0.4% gold clusters having a diameter of 25 nm. The gold clusters 2 bind by means of adsorptive forces. The probe is subsequently immersed once again in a poly(D-glucosamine) solution such that the gold clusters are packed into the layer sequence. For the purpose of detecting that the method is working, the probe is dipped to half its extent into PAA. In this way, a section of the probe is coated with a PAA layer 7.

25

In the above-described arrangement, the label is applied to the probe for the sake of clarity. In practice, the label will be provided in an analogous manner on the sample.

30

Fig. 5 shows the contact of the circular probe with the coated aluminum oxide substrate 5. In the figure, it is possible to discern a boundary line running from the top left to the bottom right. The light region corresponds to the region of the probe which is coated with a poly(D-glucosamine) layer 8. The dark region

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corresponds to the section of the probe which carries the PAA layer 7 on its surface. The repulsive interaction between the superficial PAA layer 7 on the probe and the PAA layer 7 on the substrate results in a light reflection which is different from that in the contact region of the poly(D-glucosamine) layer 8 on the probe, which layer interacts attractively with the PAA layer 7 on the aluminum substrate 5. It is consequently possible to distinguish clearly whether a label which is formed, for example, from PAA or another polymer, in particular a biopolymer, and which is applied to the aluminum substrate 5 has affinity to the probe.

## Patent claims

1. A method for identifying a first polymer (4, 7)  
which is bound to a first phase (5) which reflects  
5 electromagnetic waves, which method has the  
following steps:
  - a) bringing the first polymer (4, 7) into contact  
to a second polymer (3, 8) which has affinity to  
10 the first polymer (4, 7) and which is bound, by  
way of metallic clusters (2), to a solid second  
phase (1) which is pervious to electromagnetic  
waves,
  - 15 b) irradiating the second phase (1) with  
electromagnetic waves, and
  - c) detecting the change in the properties of the  
reflected electromagnetic waves.
- 20 2. The method as claimed in claim 1, characterized in  
that light, preferably LASER light, is used as the  
electromagnetic waves.
- 25 3. The method as claimed in either of the preceding  
claims, characterized in that the property change  
which is measured is the absorption in a  
predetermined spectrum before and/or after the  
first polymer (4, 7) has been brought into contact  
30 with the second polymer (3, 8).
4. The method as claimed in one of the preceding  
claims, characterized in that, when monochromatic  
light is used, the property change which is  
35 measured is the spectral shift.
5. The method as claimed in one of the preceding  
claims, characterized in that the property change

which is measured is the chronological change in the absorption and/or reflection when the first polymer (4, 7) and the second polymer (3, 8) are brought into contact and/or separated.

5

6. The method as claimed in one of the preceding claims, characterized in that the property change is measured at several angles of incidence which differ from each other.

10

7. The method as claimed in one of the preceding claims, characterized in that the metallic clusters (2) are evaporation-coated directly onto the second phase (1) or are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).

15

8. The method as claimed in claim 7, characterized in that a layer formed from the second polymer (3, 8) is applied to the surface of the second phase (1).

20

9. The method as claimed in claim 8, characterized in that at least one layer formed from the first polymer (4, 7) is intercalated between the layer which is applied to the surface and the layer which is bonded to the metallic clusters (2).

25

10. The method as claimed in one of the preceding claims, characterized in that a layer formed from the first polymer (4, 7) is applied to the surface of the first phase (5).

30

11. The method as claimed in claim 10, characterized in that a layer sequence formed from the first polymer (4, 7) and the second polymer (3, 8) is applied to the surface.

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12. The method as claimed in one of the preceding claims, characterized in that the first polymer (4, 7) and/or second polymer (3, 8) employ and/or employs DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptide nucleic acid (PNA) or a ligand thereof, or polyacrylic acid, polyethylenimine or poly(D-glucosamine).
13. The method as claimed in one of the preceding claims, characterized in that, in the step denoted with the letter a, at least one other polymer, which is bound to the first phase (5), is brought into contact with the second polymer (3, 8).
14. The method as claimed in claim 13, characterized in that the polymers (3, 4, 7, 8) are applied to the first phase (5) in the form of a barcode.
15. A device for identifying a first polymer (4, 7) which is bound to a first phase (5) which reflects electromagnetic waves, characterized in that a second polymer (3, 8), which has affinity to the first polymer (4, 7), is bound, by way of metallic clusters (2), to the surface of a second phase (1) which is pervious to electromagnetic waves.
16. The device as claimed in claim 15, characterized in that the metallic clusters (2) are formed from silver, gold, aluminum, copper or indium.
17. The device as claimed in claim 15 or 16, characterized in that the electromagnetic waves are light, preferably LASER light.
18. The device as claimed in one of claims 15 to 17, characterized in that the second phase (1) is produced from a transparent material, such as plastic or glass.



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19. The device as claimed in one of claims 15 to 18, characterized in that the first polymer (4, 7) and/or the second polymer (3, 8) is/are DNA, RNA, ssDNA or ssRNA or synthetic analogs thereof, protein, peptide, peptidenucleic acid (PNA) or a ligand thereof, or polyacrylic acid, poly(D-glucosamine) or polyethylenimine.
20. The device as claimed in one of claims 15 to 19, characterized in that a contrivance for determining the optical property of the reflected light is provided.
21. The device as claimed in claim 20, characterized in that the contrivance can be used to measure the absorption in a predetermined spectrum before and/or after the first polymer (4, 7) and the second polymer (3, 8) are brought into contact.
22. The device as claimed in claim 20 or 21, characterized in that the contrivance can be used to measure the spectral shift of the reflective light.
23. The device as claimed in one of claims 15 to 23, characterized in that the contrivance can be used to measure the optical property at several angles of incidence which differ from each other.
24. The device as claimed in one of claims 15 to 23, characterized in that the metallic clusters (2) are bound to the second phase (1) by way of a layer which is formed from the second polymer (3, 8).
25. The device as claimed in claim 24, characterized in that a layer which is formed from the second

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polymer (3, 8) is applied to the surface on the second phase (1).

- 5 26. The device as claimed in claim 25, characterized in that at least one layer which is formed from the first polymer (4, 7) is intercalated between the layer provided on the surface and the layer which is bonded to the metallic clusters (2).
- 10 27. The device as claimed in one of claims 15 to 26, characterized in that a layer which is formed from the first polymer (4, 7) is applied to the surface of the first phase (5).
- 15 28. The device as claimed in claim 27, characterized in that a layer which is formed from the second polymer (3, 8) is applied to the layer which is provided on the surface.

(12) NACH DEM VERTRAG ÜBER DIE INTERNATIONALE ZUSAMMENARBEIT AUF DEM GEBIET DES  
PATENTWESENS (PCT) VERÖFFENTLICHTE INTERNATIONALE ANMELDUNG

(19) Weltorganisation für geistiges Eigentum  
Internationales Büro



(43) Internationales Veröffentlichungsdatum  
21. Dezember 2000 (21.12.2000)

PCT

(10) Internationale Veröffentlichungsnummer  
**WO 00/77496 A1**

(51) Internationale Patentklassifikation<sup>7</sup>: **G01N 21/75,**  
33/543, 21/63

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(21) Internationales Aktenzeichen: PCT/DE00/01945

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(22) Internationales Anmeldedatum:  
10. Juni 2000 (10.06.2000)

(81) Bestimmungsstaaten (national): CA, JP, KR, US.

(25) Einreichungssprache: Deutsch

(84) Bestimmungsstaaten (regional): europäisches Patent (AT,  
BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC,  
NL, PT, SE).

(26) Veröffentlichungssprache: Deutsch

(30) Angaben zur Priorität:  
199 27 051.1 14. Juni 1999 (14.06.1999) DE

**Veröffentlicht:**

- Mit internationalem Recherchenbericht.
- Vor Ablauf der für Änderungen der Ansprüche geltenden  
Frist; Veröffentlichung wird wiederholt, falls Änderungen  
eintreffen.

(71) Anmelder (für alle Bestimmungsstaaten mit Ausnahme  
von US): NOVEMBER AKTIENGESELLSCHAFT  
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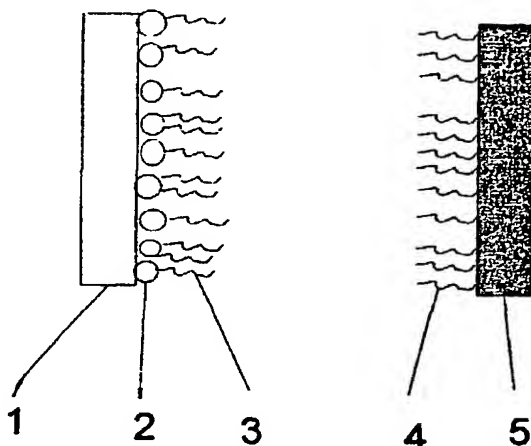
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Abkürzungen wird auf die Erklärungen ("Guidance Notes on  
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(72) Erfinder; und

(75) Erfinder/Anmelder (nur für US): BERTLING, Wolf

(54) Title: METHOD AND DEVICE FOR IDENTIFYING A POLYMER

(54) Bezeichnung: VERFAHREN UND VORRICHTUNG ZUR IDENTIFIZIERUNG EINES POLYMERS



(57) Abstract: The invention relates to a method for identifying  
a first polymer (4, 7) which is bonded to a first phase (5) that  
reflects electromagnetic waves. Said method comprises the fol-  
lowing steps: (a) bringing the first polymer (4, 7) into contact  
with a second polymer (3, 8), which is bonded to a solid second  
phase (1) by metallic clusters (2), said second phase being per-  
meable to electromagnetic waves; (b) irradiating the second phase  
(1) with electromagnetic waves; and (c) determining the change  
in the properties of the reflected electromagnetic waves.

(57) Zusammenfassung: Die Erfindung betrifft ein Verfahren  
zur Identifizierung eines an einer elektromagnetischen Wellen  
reflektierenden ersten Phase (5) gebundenen ersten Polymers  
(4, 7) mit folgenden Schritten: (a) Inkontaktbringen des ersten  
Polymers (4, 7) mit einem zweiten Polymer (3, 8), das über  
metallische Cluster (2) an eine feste für elektromagnetische  
Wellen durchlässige zweite Phase (1) gebunden ist, (b)  
Durchstrahlen der zweiten Phase (1) mit elektromagnetischen  
Wellen und, (c) Erfassen der Änderung der Eigenschaften der  
reflektierten elektromagnetischen Wellen.

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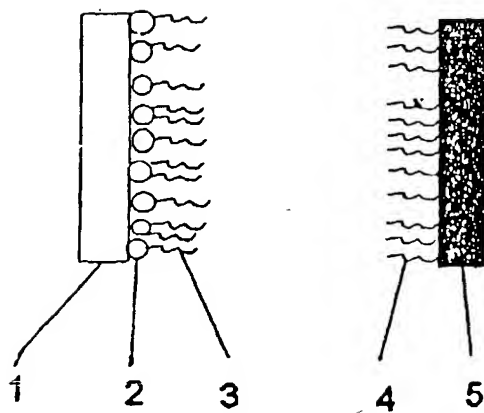


Fig. 1

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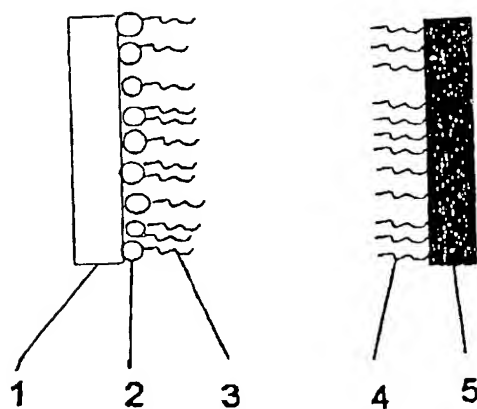


Fig. 1

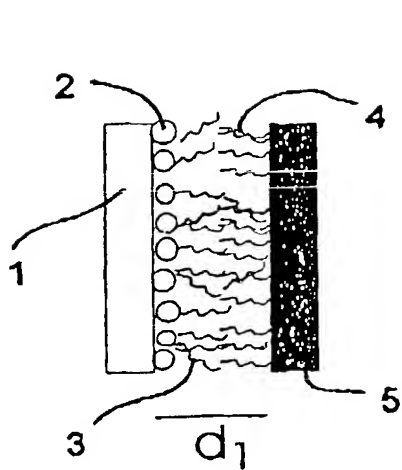


Fig. 2

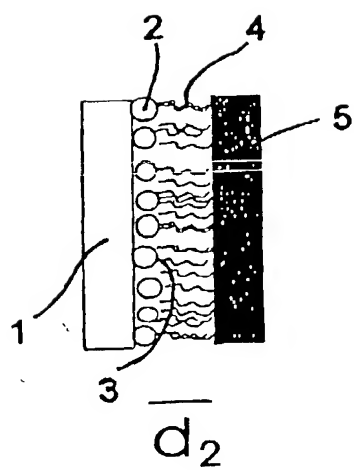


Fig. 3

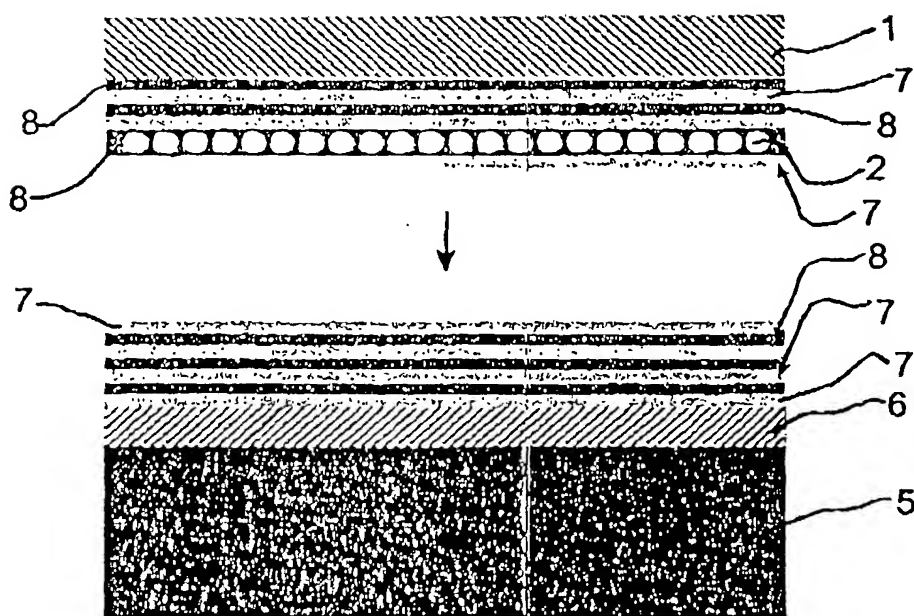


Fig. 4

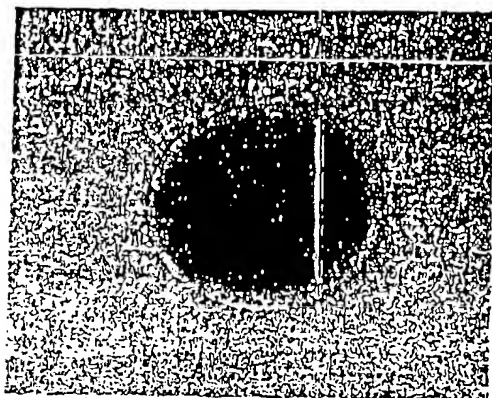


Fig 5

## COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled *Method and Device For Identifying a Polymer*, the specification of which:

- ☐ is attached hereto.  
☒ was filed on December 14, 2001 as Application Serial No. 10/018,449.  
☐ was described and claimed in PCT International Application No. \_\_\_\_\_ filed on \_\_\_\_\_ and as amended under PCT Article 19 on \_\_\_\_\_.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

Country	Application No.	Filing Date	Priority Claimed
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Germany	199 27 051.1	June 14, 1999	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No

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